

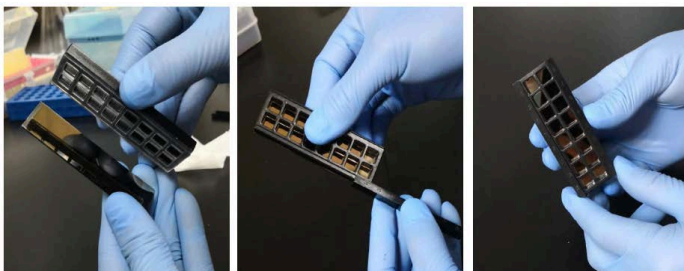
SARS-CoV-2 Protein Microarray (2-in-1 Protein/Peptide Array)

Application Notes:

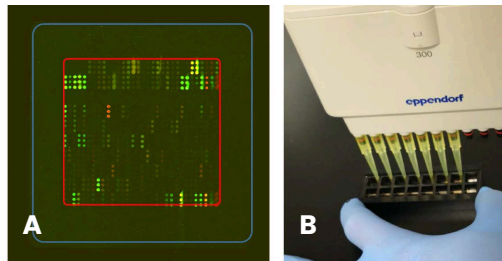
- Total *E. coli* Lysate is needed for serum profiling to prevent potential non-specific binding of the antibodies in serum against potential bacterial components, as protein contained in this array was produced in a *E. coli* expression system.
- The gasket can be reused. It should be thoroughly washed using detergent followed by clean water and air-dried after use.
- In steps 4-6, the slides should be shielded from light.
- Materials needed (not provided)
 1. ProPlate® Multi-Well Slide Module (gracebio.com/product/pro-plate-multi-well-chambers-244864)
 2. *E. coli* lysate (e.g. BioRad *E. coli* protein lysate 1632110 (www.bio-rad.com/en-us/product/e-coli-lysate-sample?ID=0e1226fb-c003-479c-8f25-057df2c7eff4).
 3. Plastic 4-well plates to store the protein microarrays during the blocking, reaction and washing steps (e.g. Thermo Scientific *Nunc* Dishes, Rectangular 4-Well, No.12-565-495), or use a larger container
 4. Aluminum foil.

Protocol for Serum Profiling:

1. **Blocking.** The SARS-CoV-2 protein microarrays have been blocked prior to shipment.
2. **Incubation of sera.** Place the 16-chamber rubber gasket onto each slide to create individual chambers for the 14 identical sub-arrays (below) **NOTE:** The last two sub-arrays are intended to be blank.



Attach the gasket to slide and place the clips on both sides.



A. The relative size of the microarray area and the chamber. (Blue: the size of the chamber; Red: the microarray region)

B. Multi-channel pipette could be used for sample operation and liquid handling.

NOTE: Keep the pipette tips at one corner of the chamber to avoid touching the surface of the microarray region when pipetting.

Dilute sera (1:200) in PBS buffer containing 0.1% Tween 20, 1% BSA and 0.5 mg/ml total *E. coli* lysate. Then add 200 μ l of diluted serum to each of the 14 separate sub-array wells and incubate at room temperature for 2 hours.

3. **Washing.** Wash each sub-array separately three times with PBS containing 0.1% Tween 20, with a multiple-channel pipette (pumping pipette plunger 20-30 times each is recommended to help wash thoroughly) to avoid contamination among samples. Remove the gasket from the slide. Transfer the slide to a larger container and wash three times (10 min each, at a speed of 100-110 rpm) using an orbital shaker.
4. **Incubation of the secondary antibody.** Dilute secondary antibodies (according to manufactures recommended dilution) in PBS buffer containing 0.1% Tween 20 and 1% BSA and incubate at room temperature for 1 hour.

NOTE: Recommended secondary antibodies: Cy3-conjugated goat anti-human IgG (109-165-008; www.jacksonimmuno.com/catalog/products/109-165-008) and Alexa Fluor 647-conjugated donkey anti-human IgM (709-605-073); www.jacksonimmuno.com/catalog/products/709-605-073.

Place arrays in 4-well microarray plate and add secondary, then cover with aluminum foil and incubate for 1 hour with gentle shaking.

5. **Washing.** Wash three times for 5 min. with PBS containing 0.1% Tween 20 on an orbital shaker (10 min, a speed of 100-110 rpm is recommended). Wash buffer: PBS with 0.1% Tween 20. And 2 additional washes with ddH₂O (5 min. each).
6. **Dry.** Once slides are dry, they are ready to scan.